

# ***Are Viruses a Hazard in Waste Water Recharge of Urban Sandstone Aquifers?***

John Tellam, Rae Mackay, Michael  
Riley, Joanna Renshaw, Michael Rivett,  
Richard Greswell, Fernanda Aller,  
Veronique Durand

***Hydrogeology Research Group  
Earth Sciences***

***School of Geography, Earth and  
Environmental Sciences***

***University of Birmingham***

***UK***

With

Steve Pedley & Eadaoin Joyce

Robens Centre, Surrey University

***R.Mackay@bham.ac.uk***

***J.H.Tellam@bham.ac.uk***





# Aims

**To determine if viruses are a hazard  
for urban waste-water recharge  
programmes in urban sandstone  
aquifers**



# Why viruses?

For most contaminants there is at least a certain amount of experience for predicting mobility

## Viruses

comparatively little experience / data  
(worldwide)

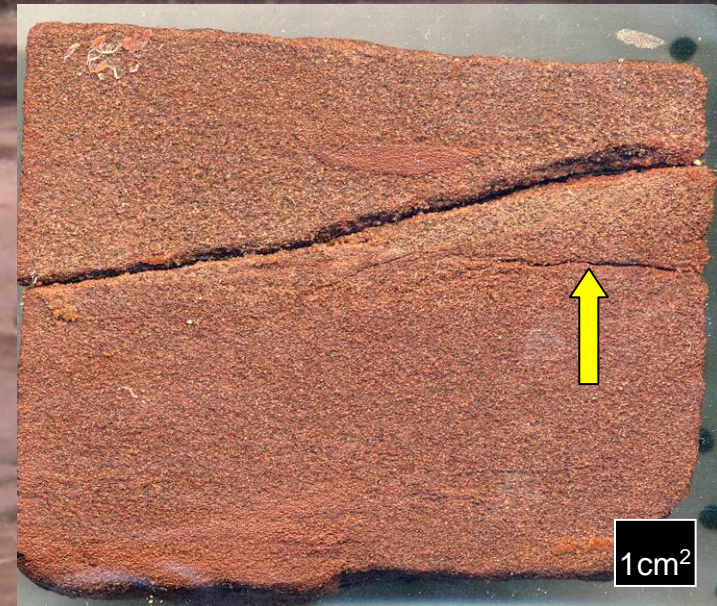
recent evidence suggests that may be an issue



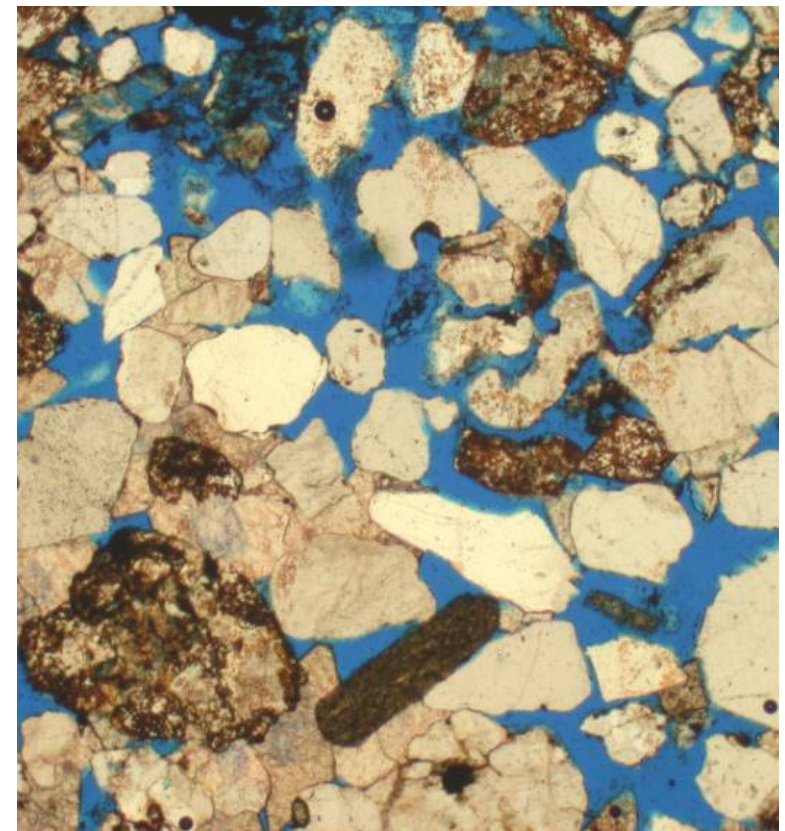
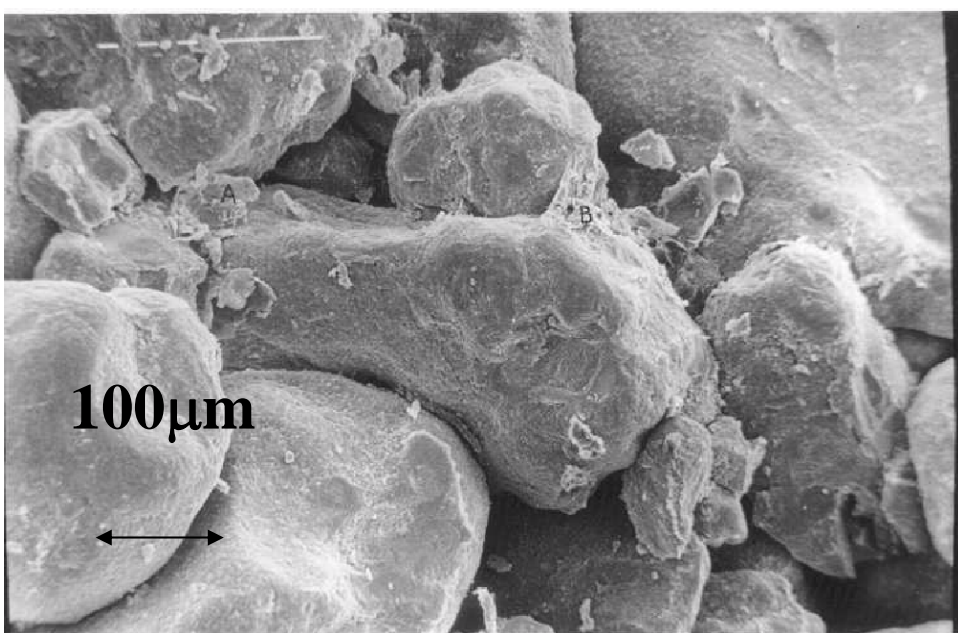
# The aquifer

Permo-Triassic Sst sequence.

UK's second most used aquifer, and equivalent red-bed sequences used across EU and beyond.







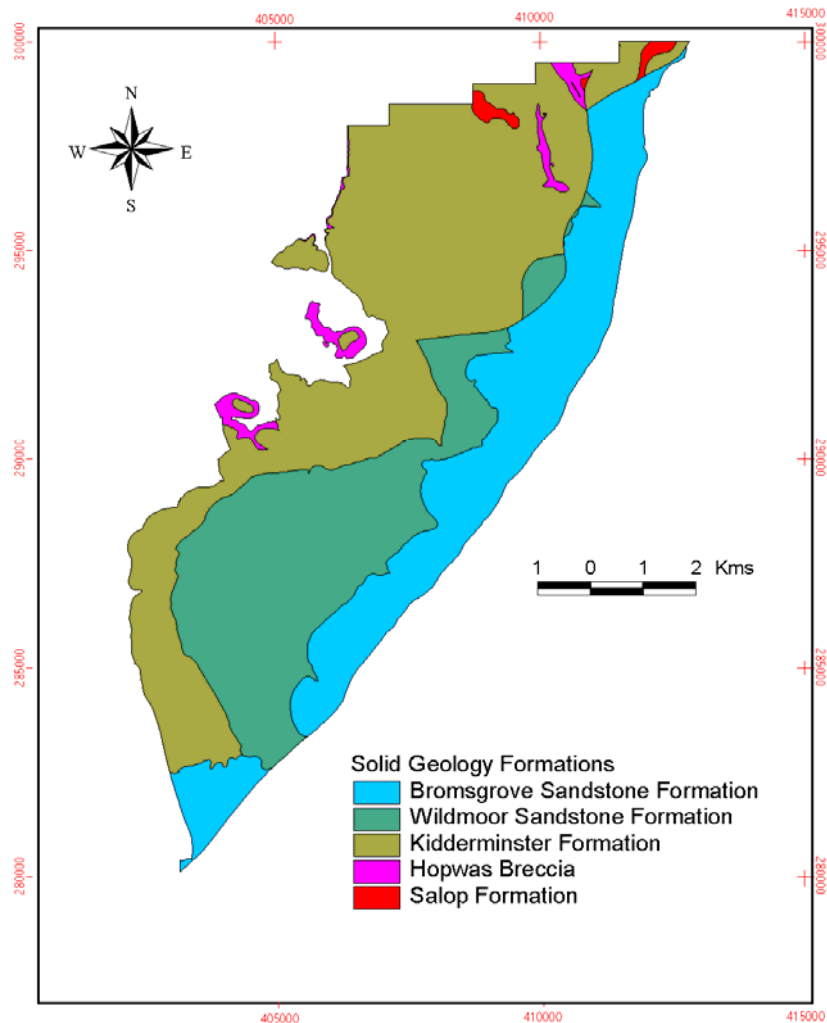
1 mm



100µm

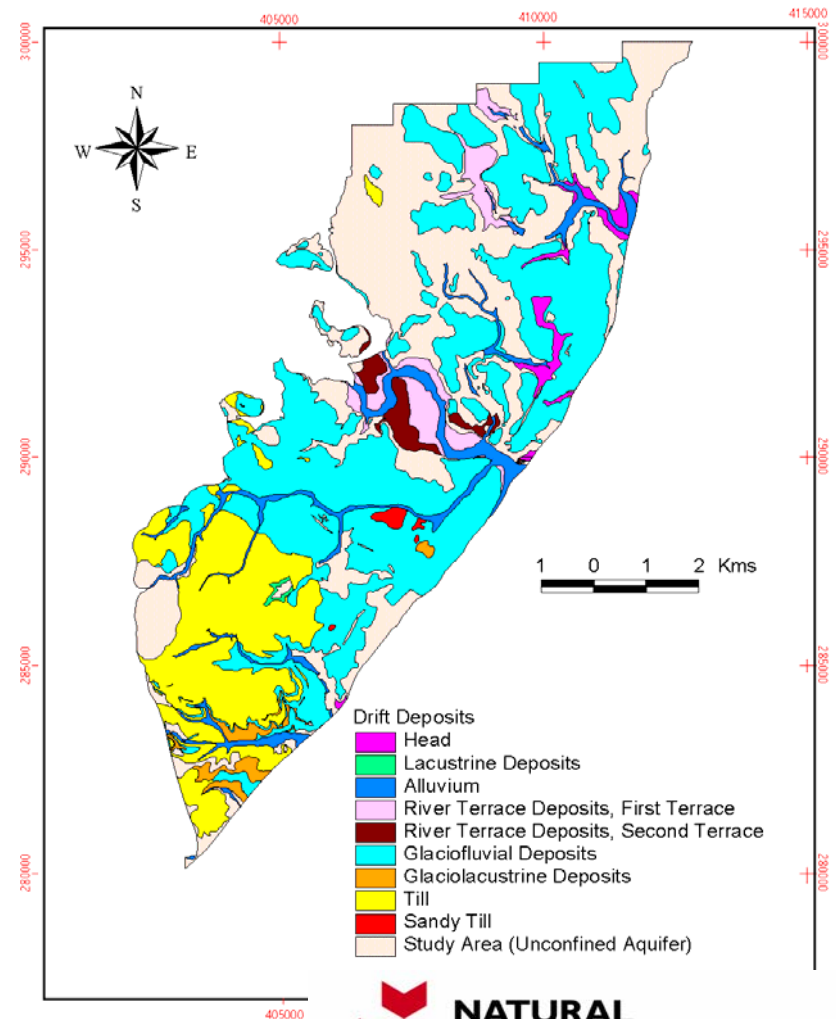
## SOLID GEOLOGY OF BIRMINGHAM AREA

(UNCONFINED AQUIFER PART ONLY)

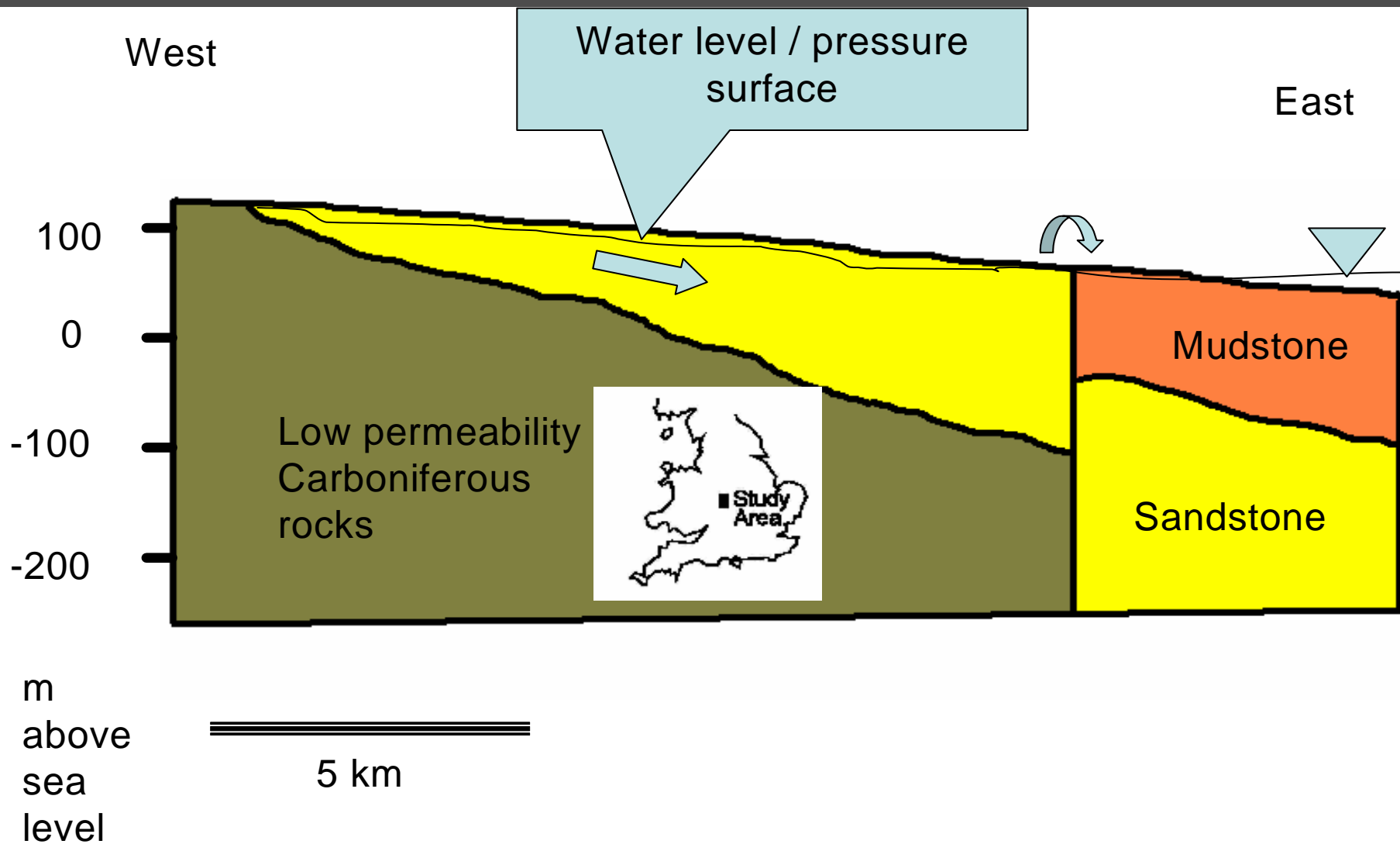


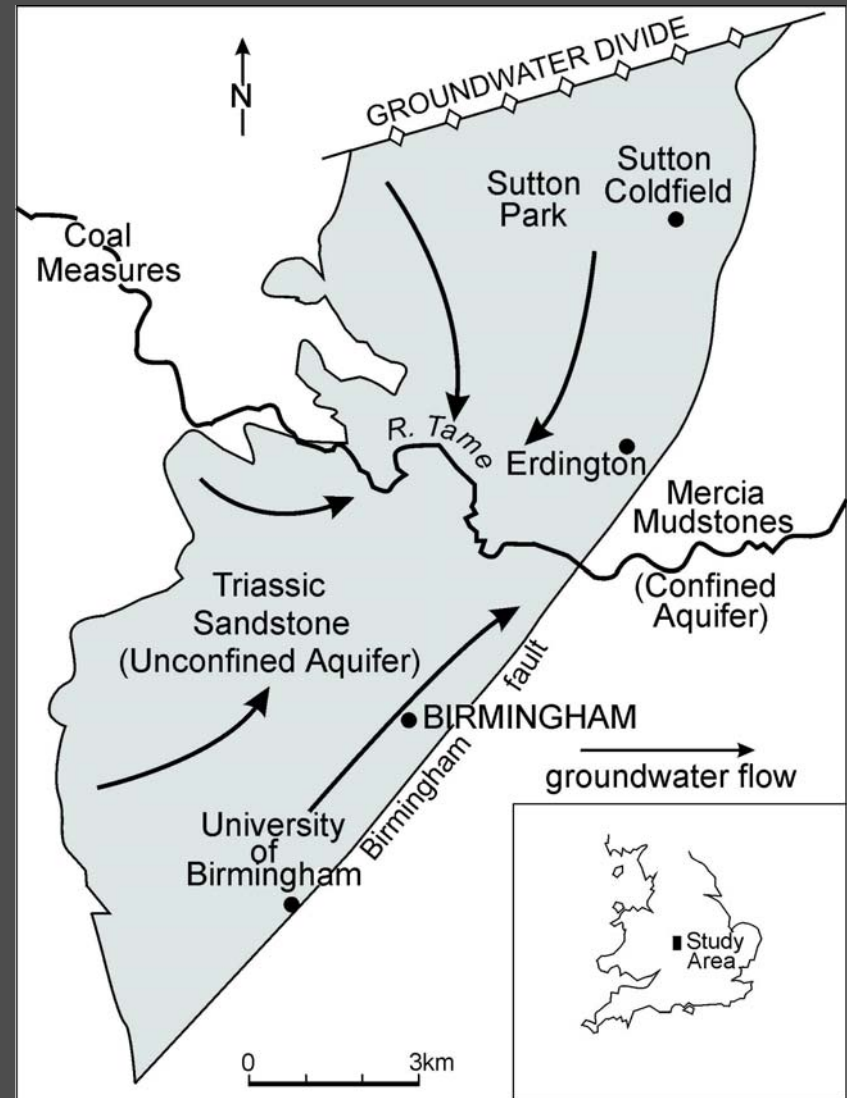
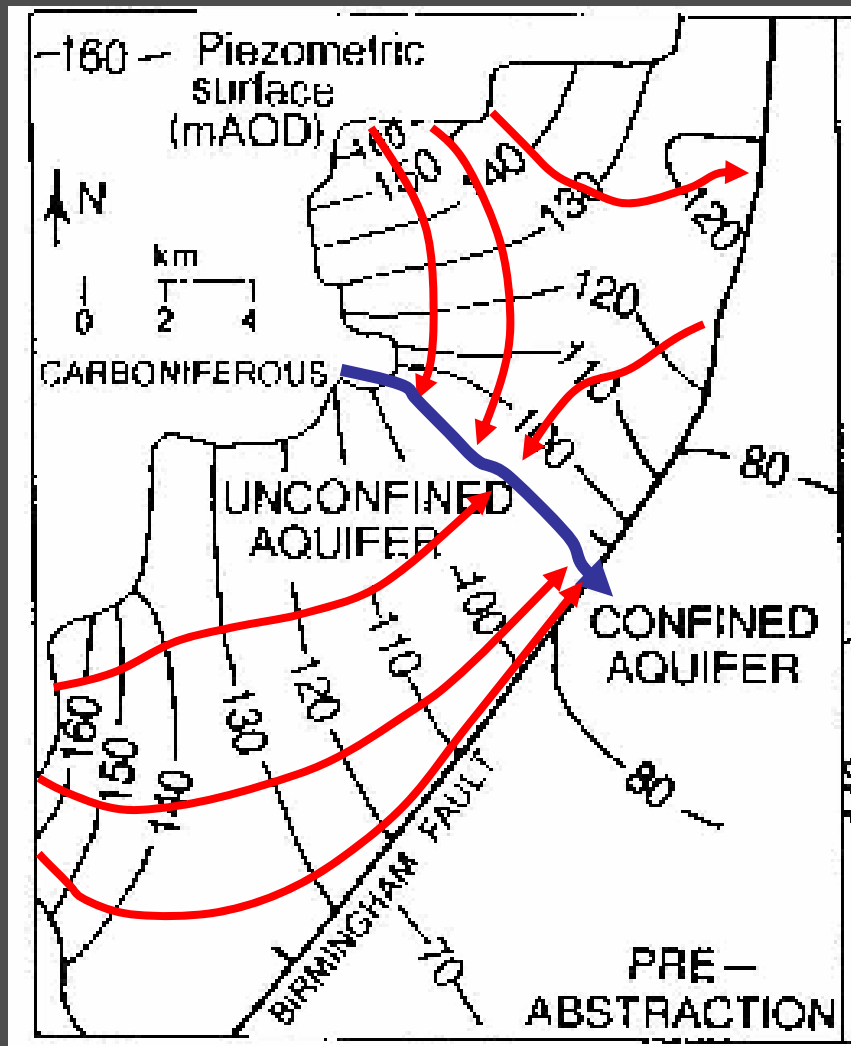
## DRIFT GEOLOGY OF BIRMINGHAM AREA

(UNCONFINED AQUIFER PART ONLY)



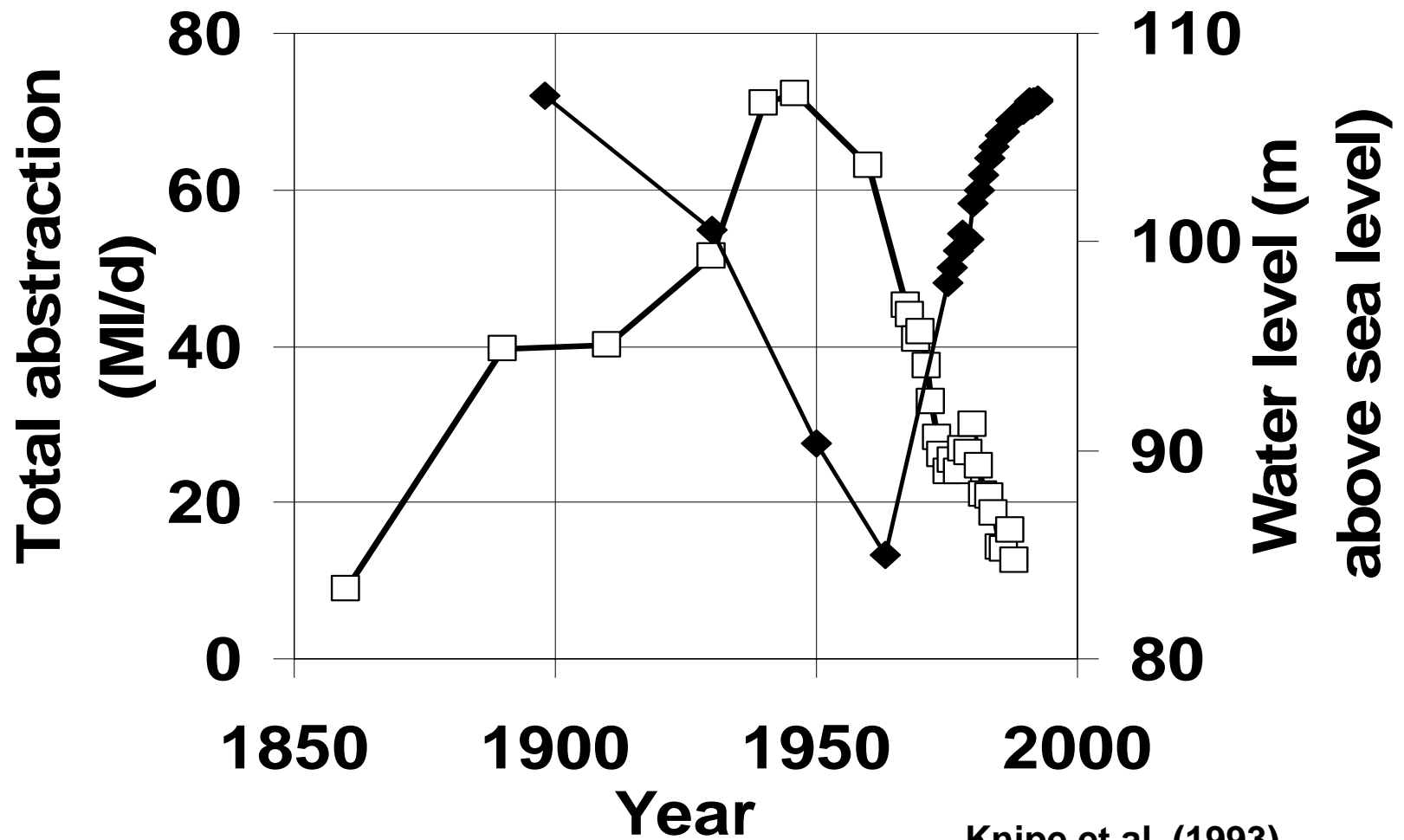
**NATURAL  
ENVIRONMENT  
RESEARCH COUNCIL**



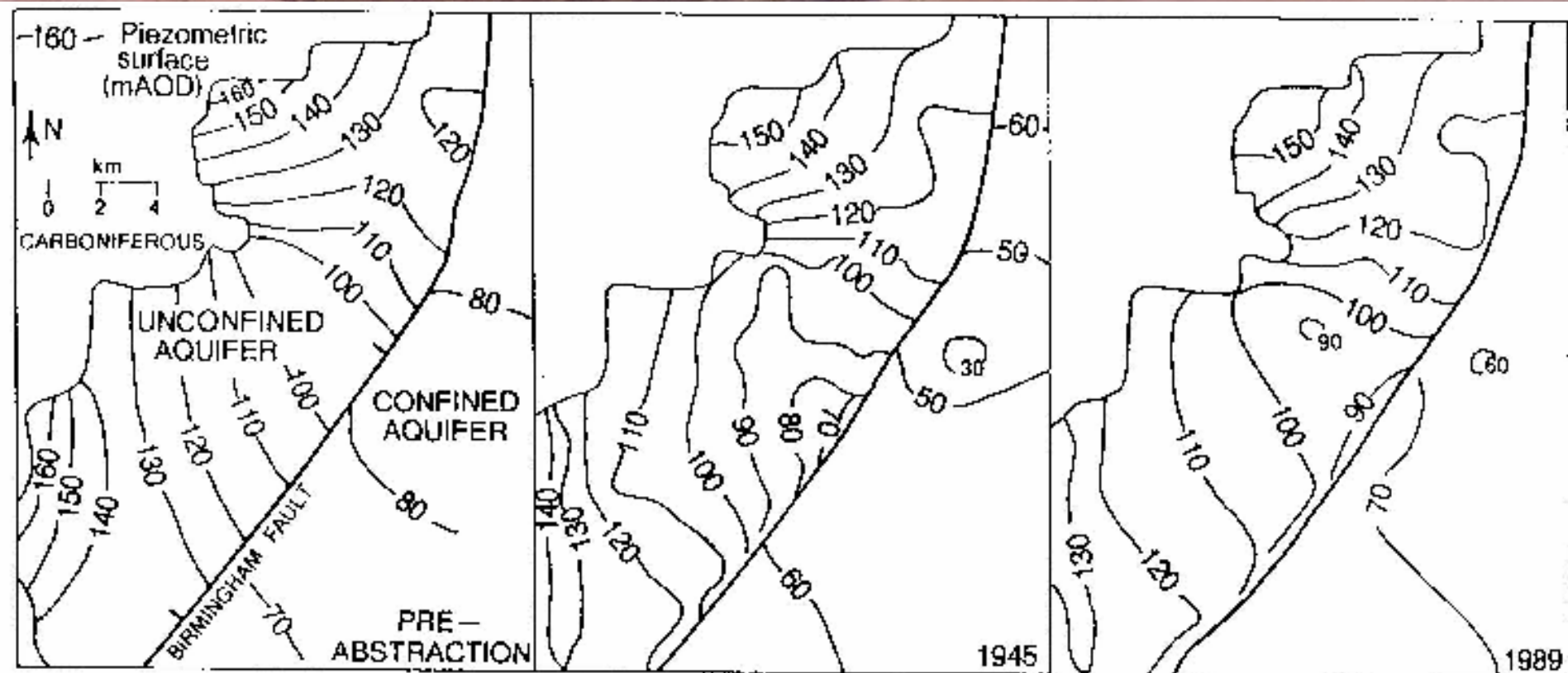


- Gw velocity ~ 10-100 m/y



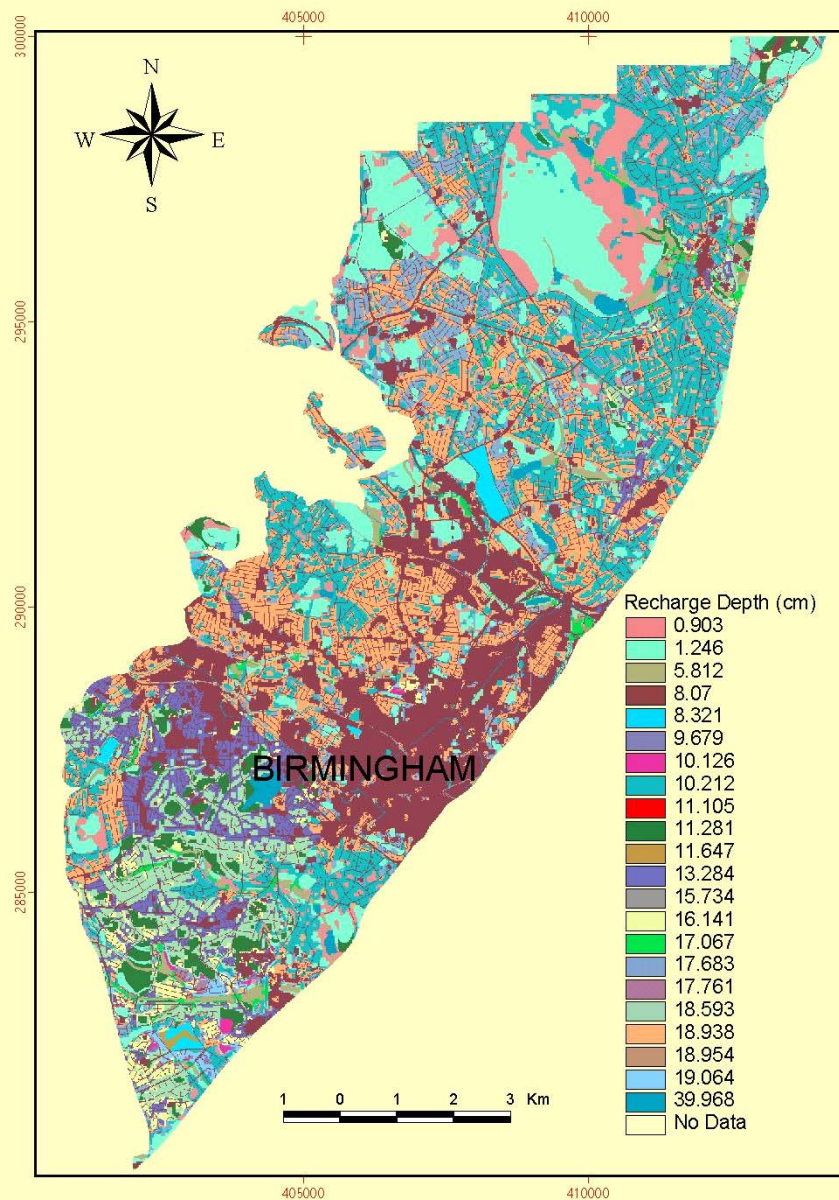


Knipe et al. (1993)



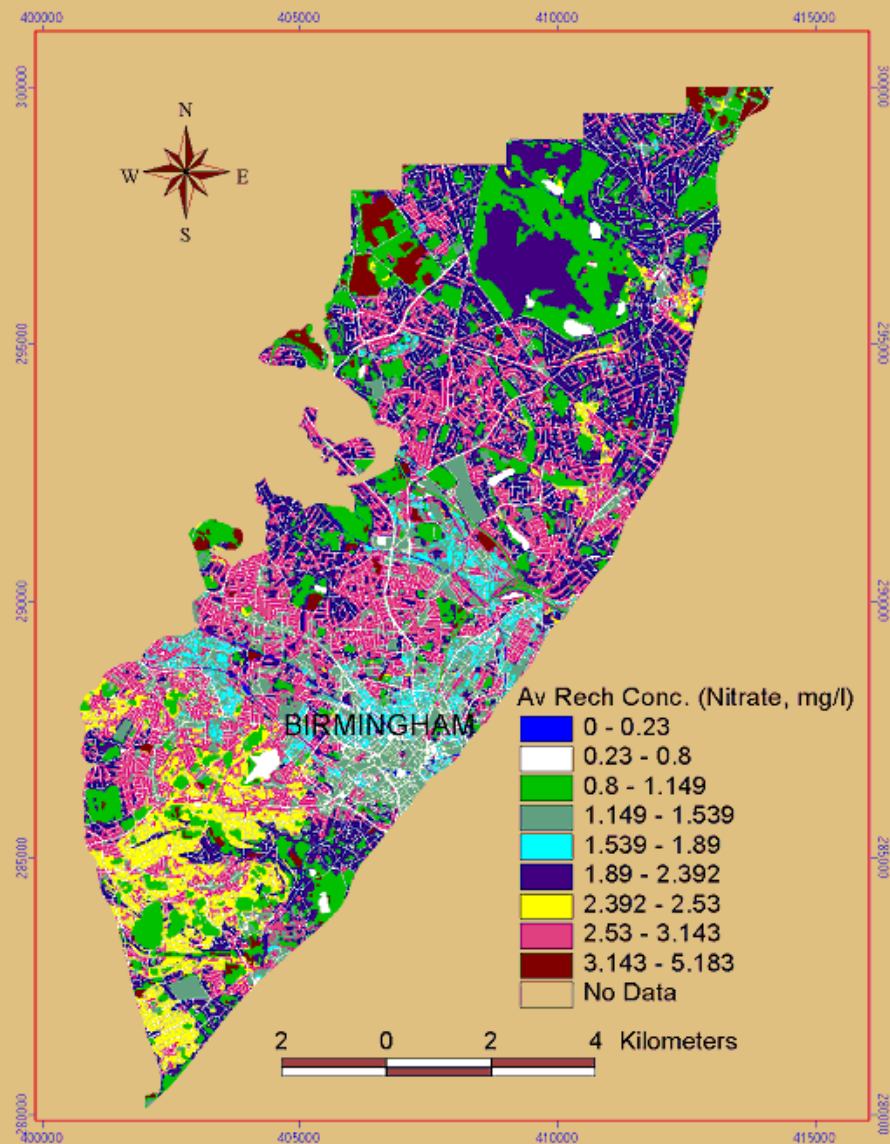


# TOTAL POTENTIAL RECHARGE DEPTHS IN BIRMINGHAM DURING WINTER (JAN - MARCH) & AUTUMN (OCT - DEC) 1980

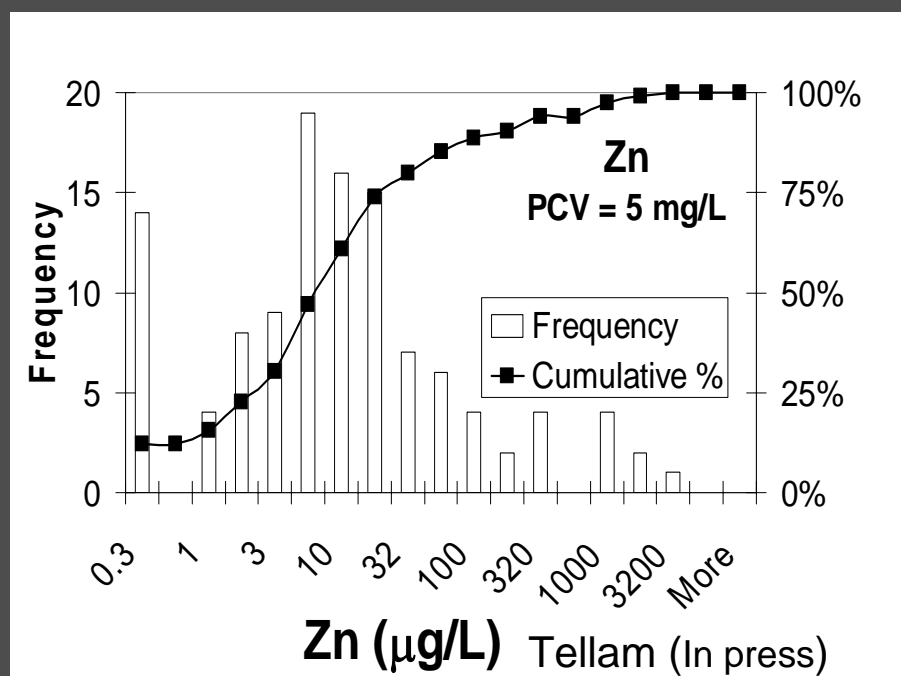
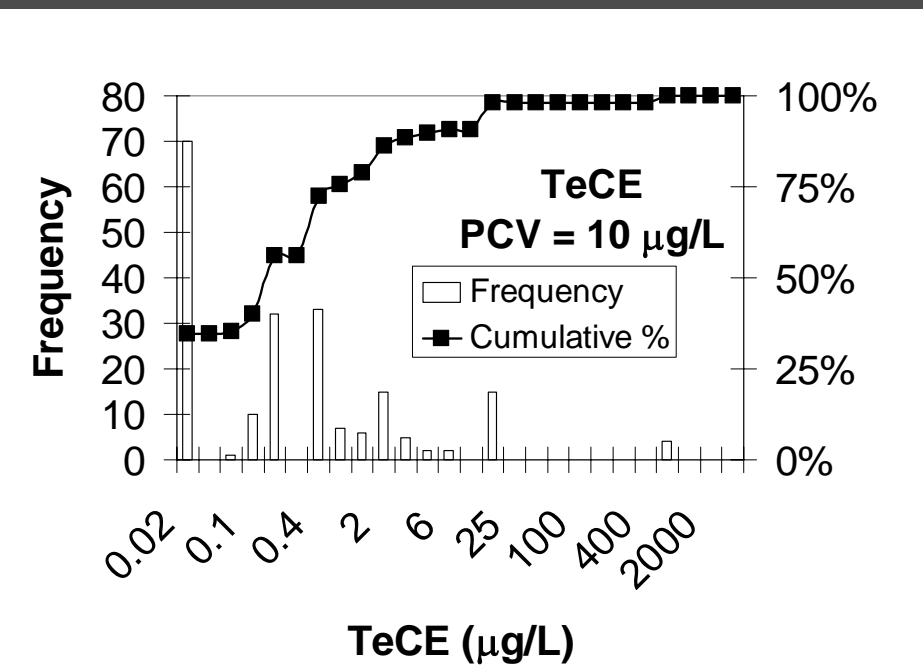
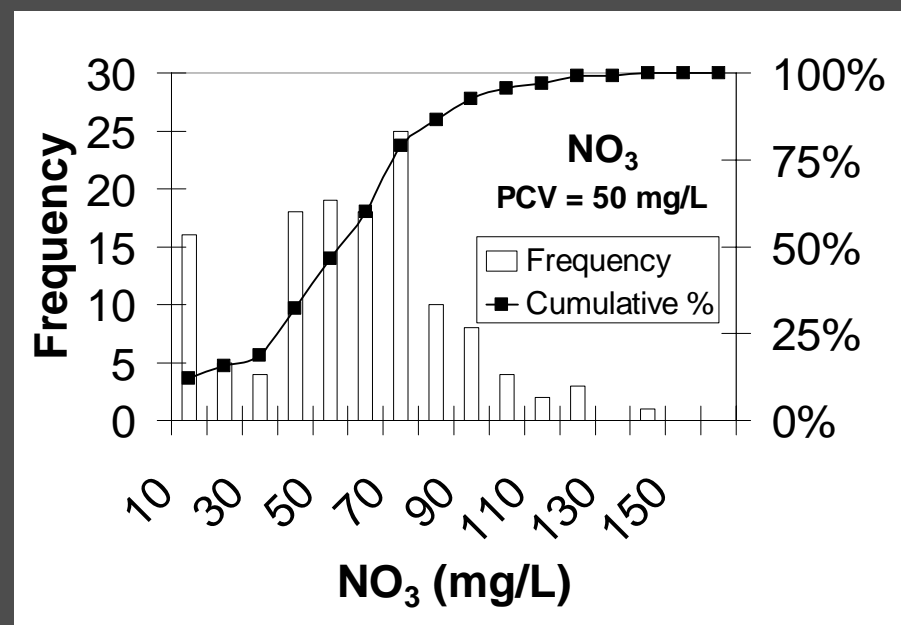
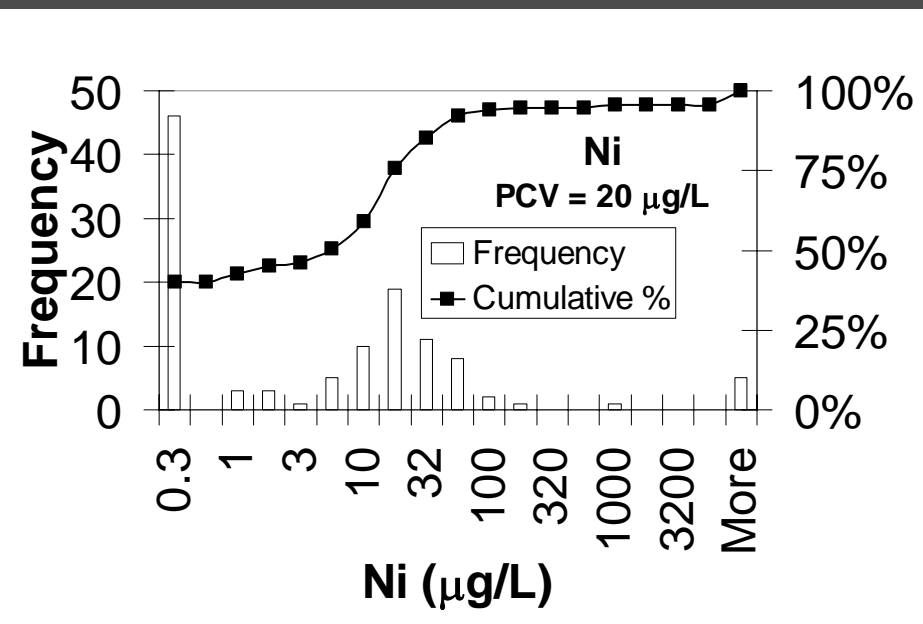


(THE MAP AREA COVERS ONLY UNCONFINED AQUIFER PART OF BIRMINGHAM)

# AVERAGE NON POINT SOURCE NITRATE CONCENTRATIONS IN RECHARGE WATERS OF BIRMINGHAM AREA DURING AUTUMN AND WINTER, 1980



THE MAP AREA COVERS ONLY UNCONFINED AQUIFER PART OF BIRMINGHAM

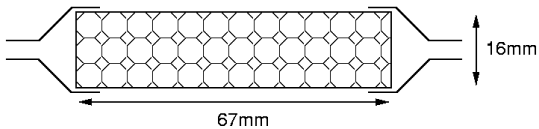




# Human virus occurrence in UK urban sand aquifers

Abstraction wells in Birmingham

EPSRC Study:  
*Enteroviruses in Groundwaters*



Date	Site	Enterovirus (PFU/L)	Rotavirus (PFU/L)
<i>Industrial Wells</i>			
19/05/99	8	2	<1
28/05/99	10	1	<1
18/06/99	10	<1	170
19/05/99	17	1	<1
11/06/99	17	<1	<1
18/06/99	B1-55K	<1	8
18/06/99	U2	<1	<1
<i>Piezos</i>			
21/04/99	B	30	450
19/05/99	B	2	<1
31/08/99	B	<1	<1
29/02/00	B	<1	-
21/04/99	D	65	<1
05/05/99	D	1	<1
31/08/99	D	<1	<1
29/02/00	D	4	-
29/02/00	E	<1	-
26/03/99	F	<1	<1
29/02/00	F	4	-



EPSRC

Piezometers in Nottingham

Powell et al. (2000)

Site A Multilevel	Depth of sample (mbgl)	Coliphage PFU/L	Enterovirus (cell culture)	Enterovirus RT-PCR	Rotavirus RT-PCR	Norwalk like viurs RT-PCR	Adenovirus PCR	Astrovirus RT-PCR
1	8.24	<1	nd	nd	nd	nd	nd	nd
2	11.22	<1	nd	nd	nd	detected	nd	nd
3	15.00	<1	nd	nd	nd	detected	nd	nd
4	18.00	<1	nd	nd	nd	nd	nd	nd
5	22.38	<1	nd	nd	nd	detected	nd	nd
6	26.45	<1	nd	nd	nd	nd	nd	nd
7	30.35	<1	nd	nd	nd	detected	nd	nd
8	35.33	30	nd	nd	nd	nd	nd	nd
9	39.30	<1	nd	nd	nd	detected	nd	nd
10	42.50	<1	nd	nd	nd	no sample	nd	nd
11	49.22	<1	nd	nd	nd	nd	nd	nd

nd - not detected

Powell et al. (2003)

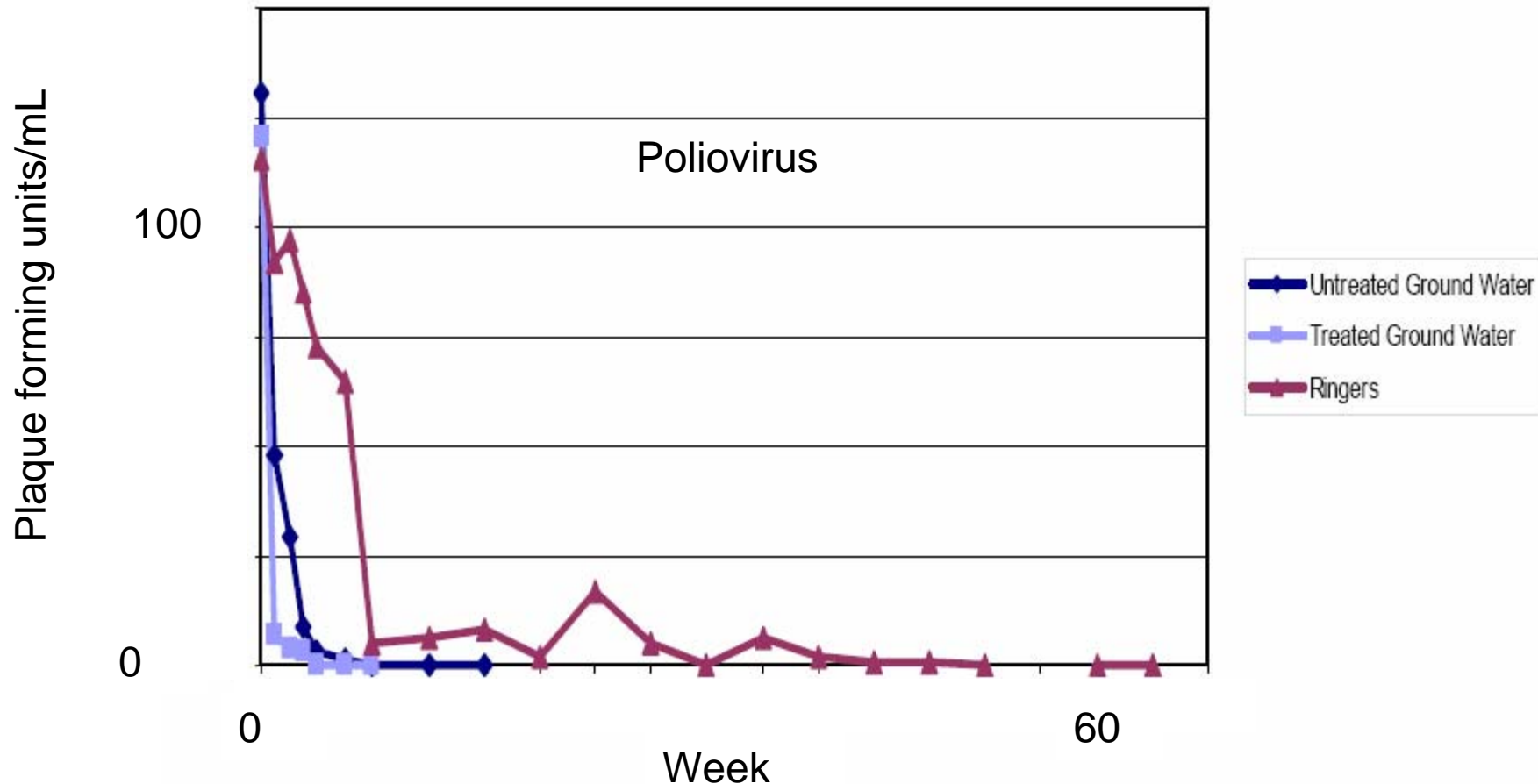


# Experimental studies of virus survival



Data from Jane Sellwood,  
Health Protection Agency

Enteric Virus Persistence in Ground Water: Study I  
Poliovirus 3 by Suspended Cell Plaque Assay



## Last detection by PCR (weeks):

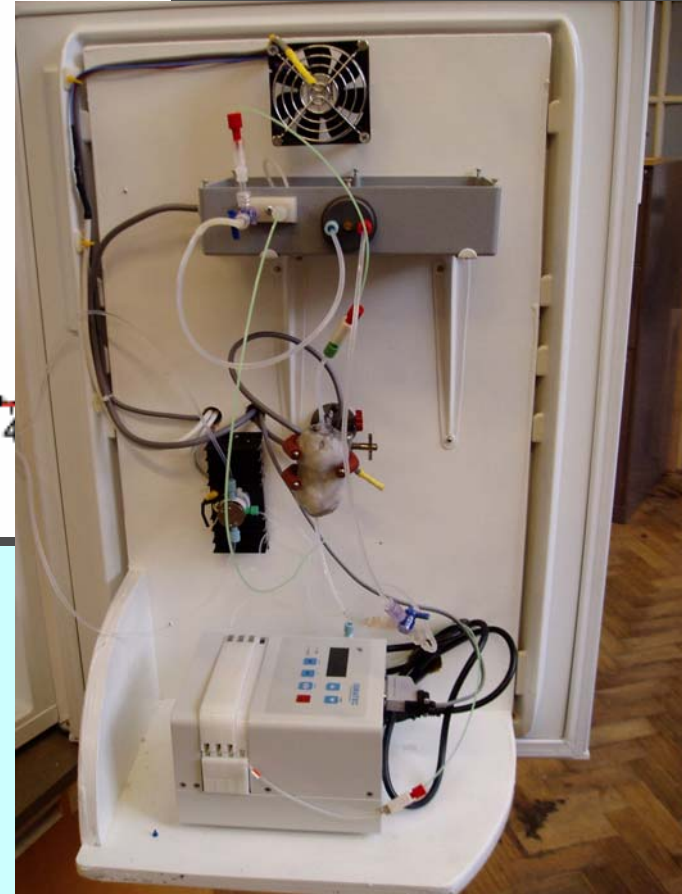
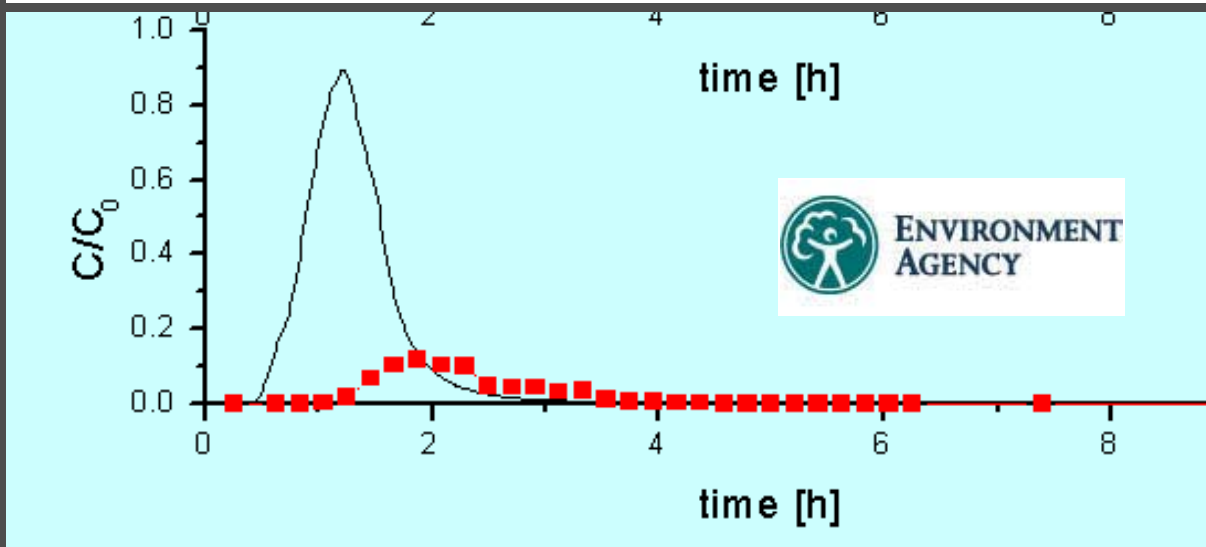
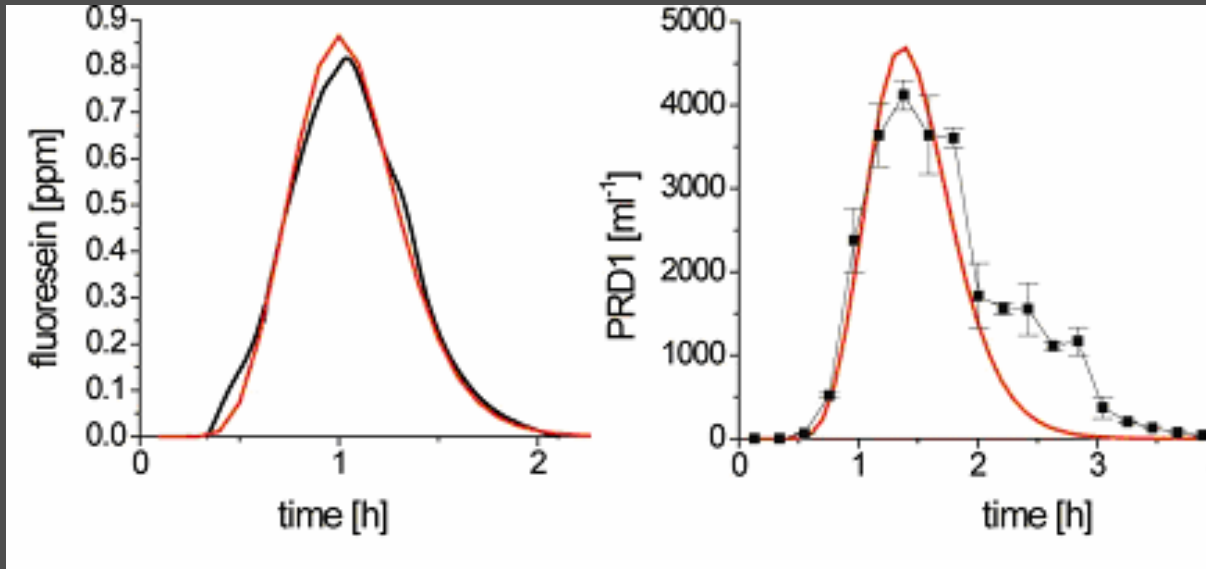
	Untreated Groundwater	Treated Groundwater	Ringers Solution
Enterovirus	32	16	>104
Adenovirus	96	>104	>104
Norovirus Genogroup I	84	100	>104
Norovirus Genogroup II	96	>104	>104

**Data from Jane Sellwood,  
Health Protection Agency**



# Experimental studies of virus mobility

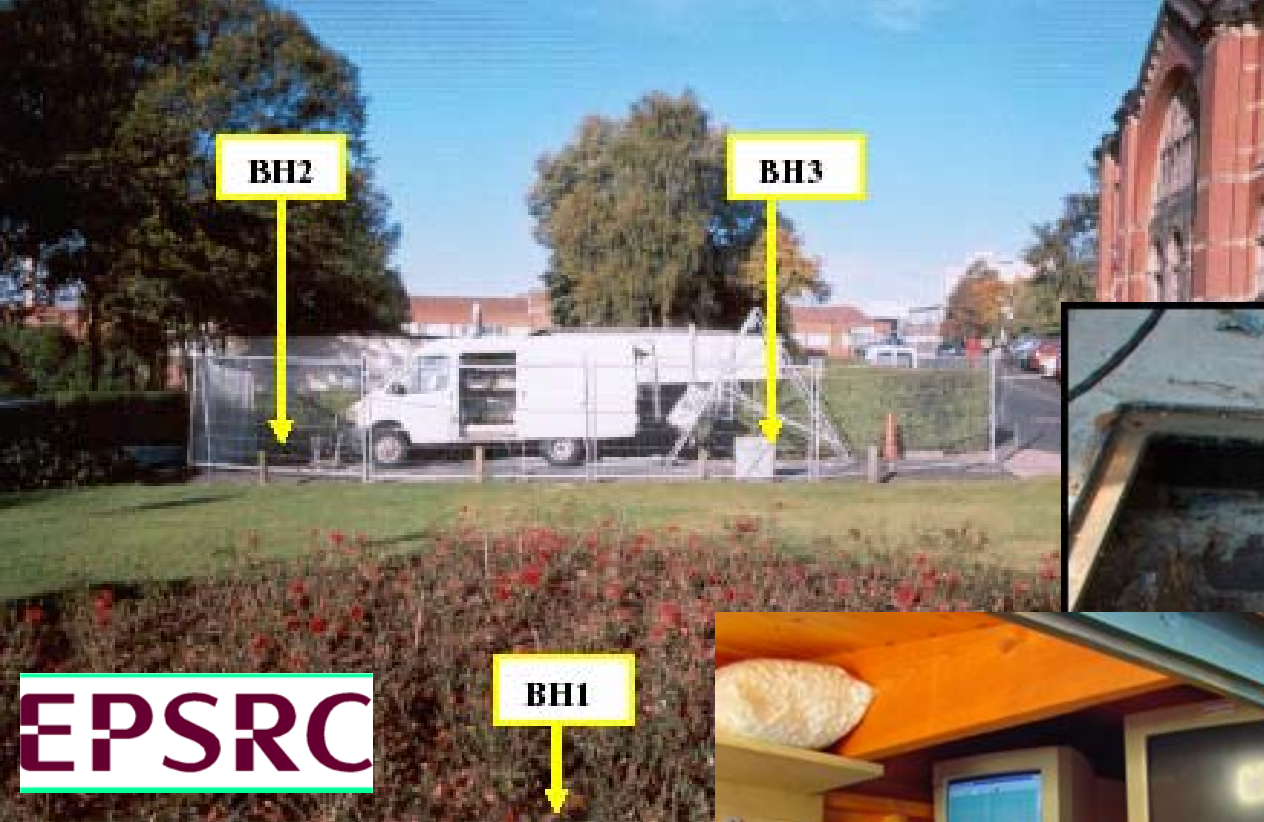
## *laboratory column expts*



ENVIRONMENT  
AGENCY

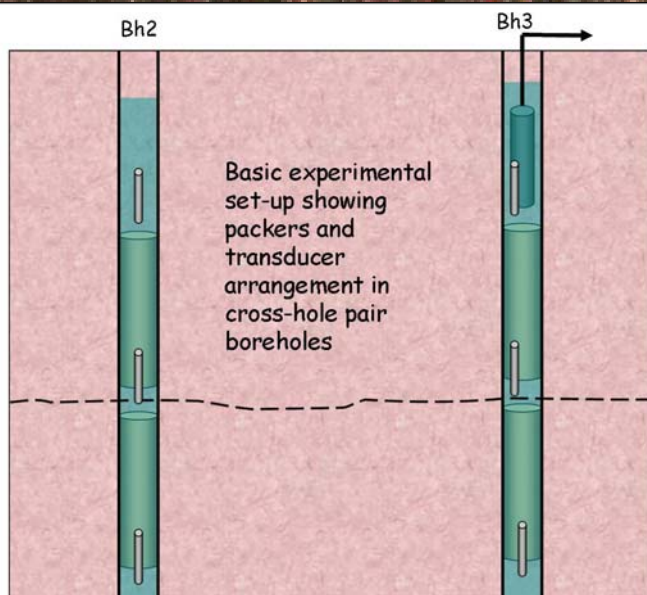


NATURAL  
ENVIRONMENT  
RESEARCH COUNCIL



*field*

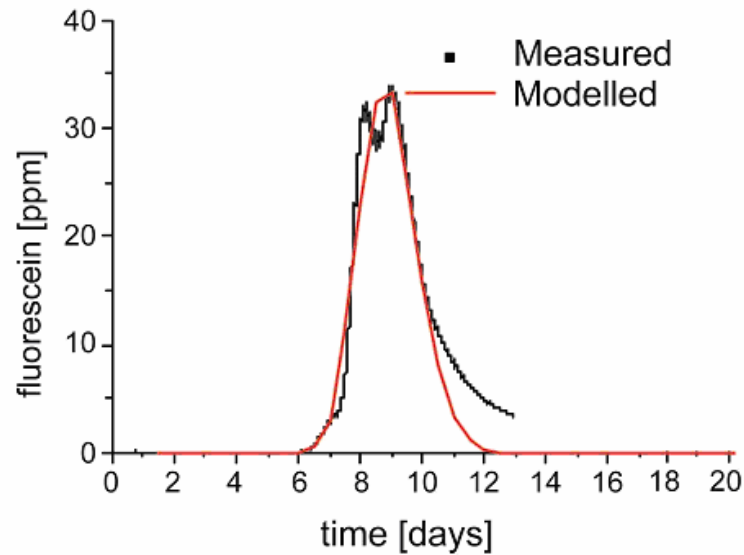
EPSRC



ENVIRONMENT  
AGENCY



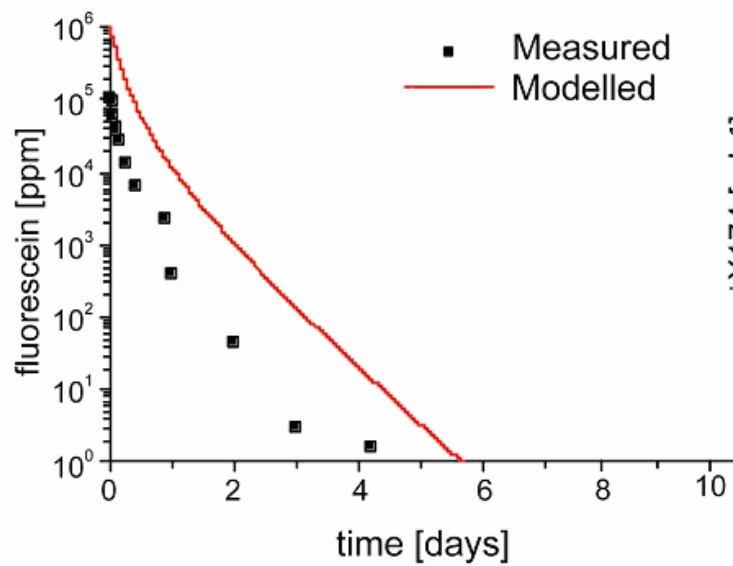
## Recovery Well



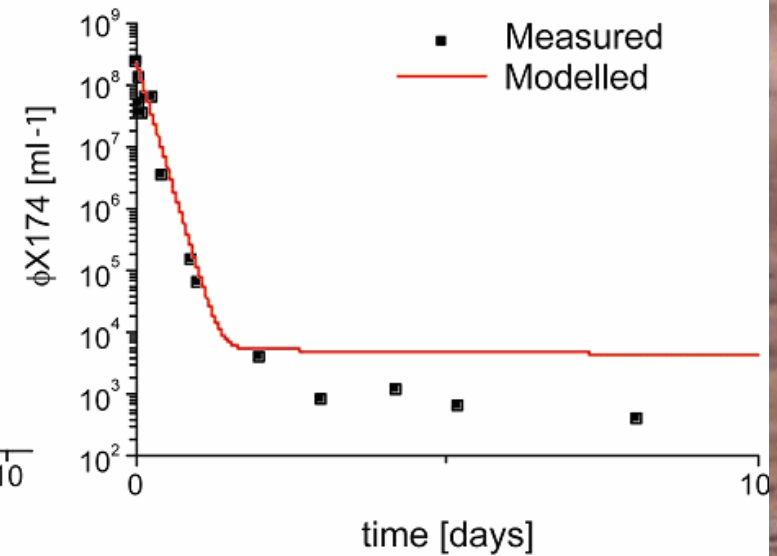
No virus  
recovered

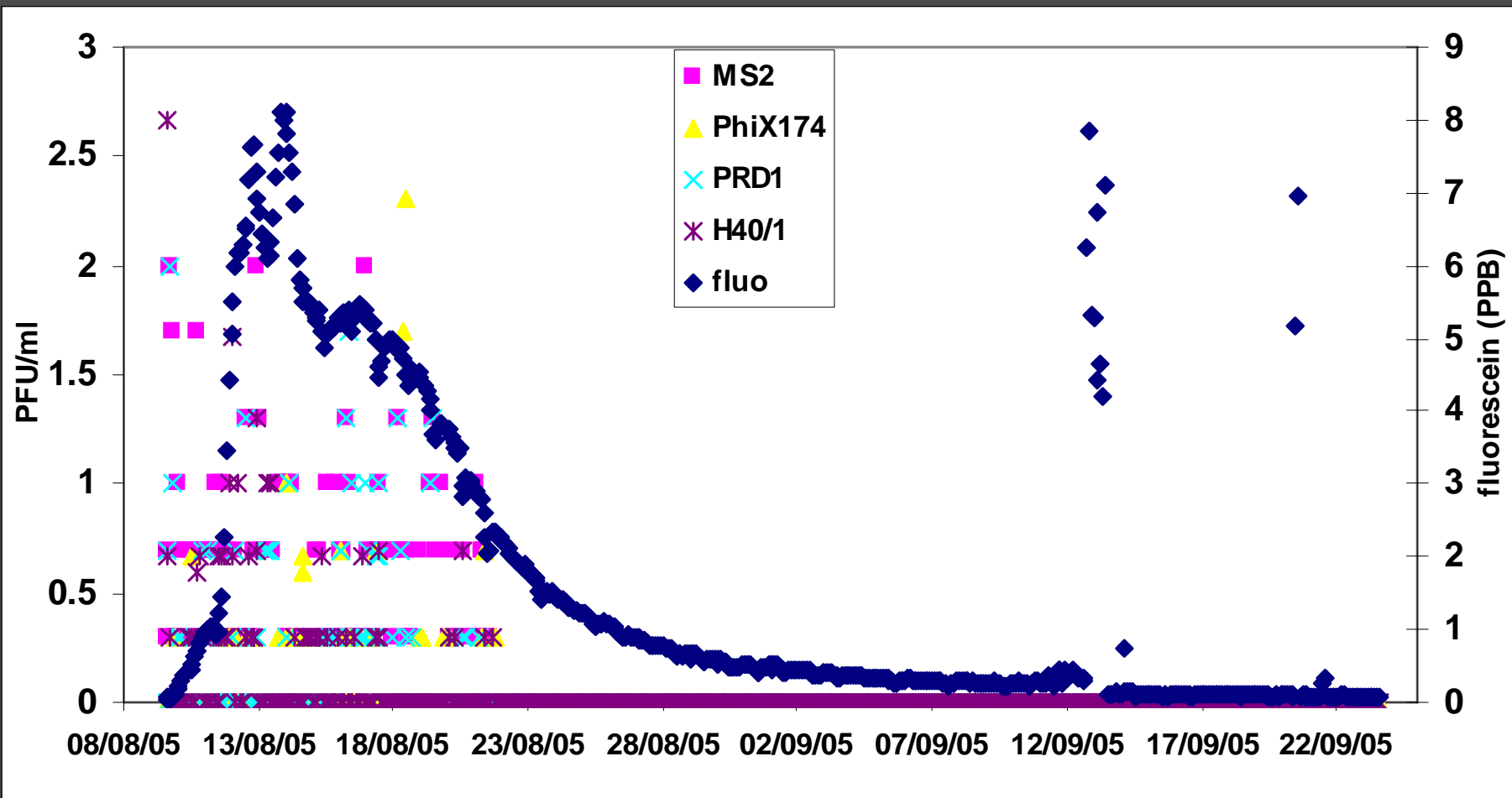
Joyce et al. (In press)

## Injection Well



## Injection Well







# The SWITCH project WP3.2: approach, issues, design, and progress

## Approach

can we use the aquifer's attenuation capacity to make sure that viruses are not present in recovered waste water?

Empirical investigation ....





The main **issues** to be addressed are:

1. velocities and distances of travel of viable viruses under aquifer conditions
2. effects of (pumping-induced) gw velocity changes
3. effect of attachment on viability



# Design and Progress

- *field monitoring*
  - velocities and distances of travel under aquifer conditions
- *field experimentation*
  - velocities and distances of travel under aquifer conditions
  - effects of pumping-induced gw velocity changes
- *laboratory experimentation*
  - effects of pumping-induced velocity changes
  - effects of attachment on viability
- *modelling*

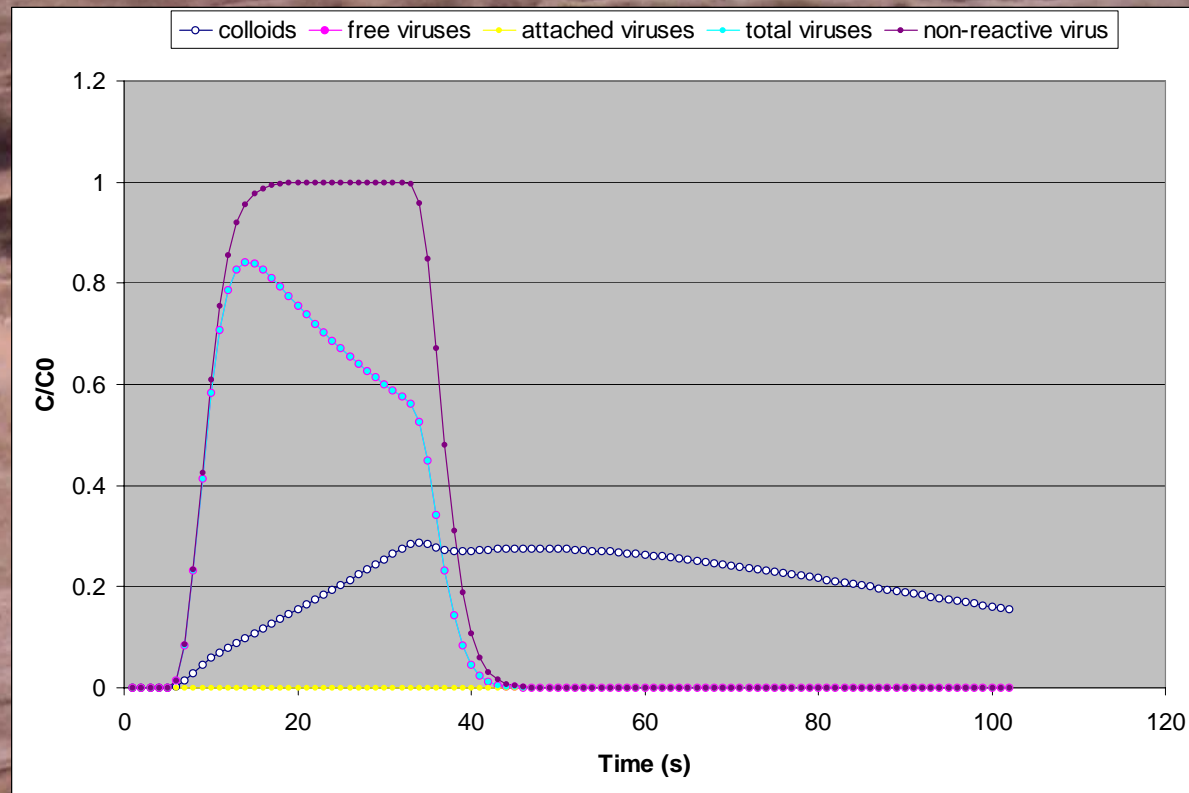
# modelling

Used in all other aspects of project:

- planning expts
- hydrogeological assessment of field systems
- interpretation of field and lab data
- extrapolation of results



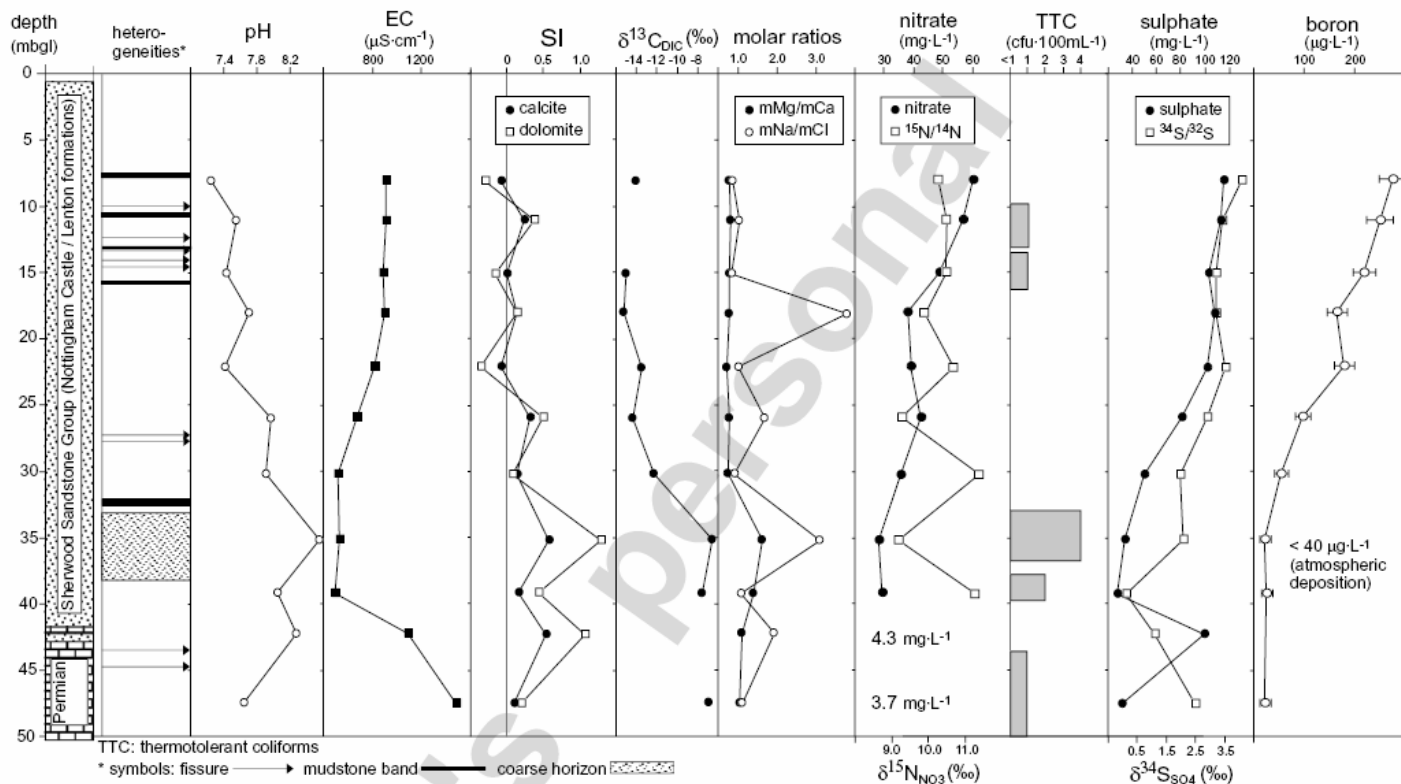
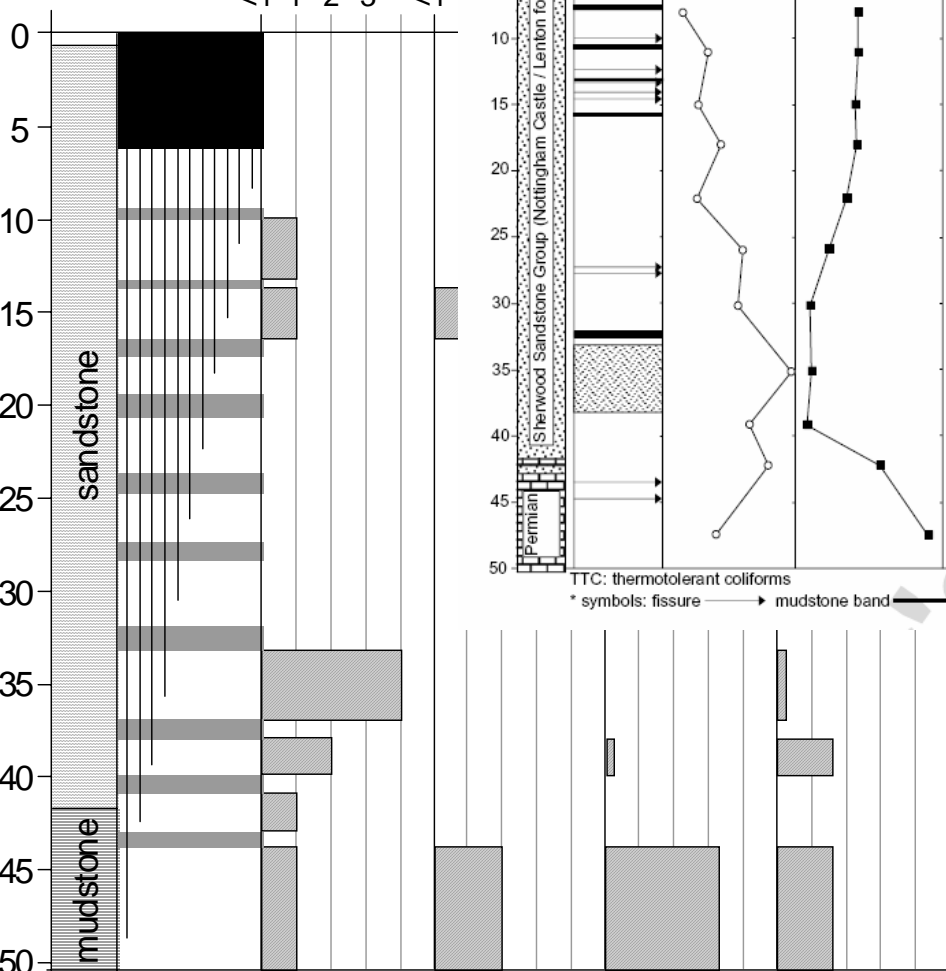
Will use flow models  
and possibly a  
colloid transport  
model developed  
in a project just  
ending





# field monitoring

Depth (mbgl)<sup>1</sup> Sampling intervals<sup>2</sup> June 6-7  
TTC (cfu/100mL)<sup>3</sup> (<1 1 2 3 <1)



- (1) mbgl: metres below ground level  
(2) sampling intervals separated by bentonite seals (gray shade)  
(3) cfu/100mL: colony-forming units / 100mL



Site A Multilevel	Depth of sample (mbgl)	Coliphage PFU/L	Enterovirus (cell culture)	Enterovirus RT-PCR	Rotavirus RT-PCR	Norwalk like viurs RT-PCR	Adenovirus PCR	Astrovirus RT-PCR
1	8.24	<1	nd	nd	nd	nd	nd	nd
2	11.22	<1	nd	nd	nd	detected	nd	nd
3	15.00	<1	nd	nd	nd	detected	nd	nd
4	18.00	<1	nd	nd	nd	nd	nd	nd
5	22.38	<1	nd	nd	nd	detected	nd	nd
6	26.45	<1	nd	nd	nd	nd	nd	nd
7	30.35	<1	nd	nd	nd	detected	nd	nd
8	35.33	30	nd	nd	nd	nd	nd	nd
9	39.30	<1	nd	nd	nd	detected	nd	nd
10	42.50	<1	nd	nd	nd	no sample	nd	nd
11	49.22	<1	nd	nd	nd	nd	nd	nd

nd - not detected

**March sampling ~ same time as clinical max NLVs: June sampling ~ Coxsackie clinical max**

Site A Multilevel	Depth of sample (mbgl)	Coliphage PFU/L	Enterovirus (cell culture) (PFU/2mL)	Enterovirus RT-PCR	Rotavirus RT-PCR	Norwalk like viurs RT-PCR	Adenovirus PCR	Astrovirus RT-PCR
1	8.26	<1	nd	nd	nd	nd	nd	nd
2	11.00	<1	1 (Coxsackie B4)	nd	nd	nd	nd	nd
3	15.22	<1	nd	nd	nd	nd	nd	nd
4	18.00	<1	1 (to be confirmed)	nd	nd	nd	nd	nd
5	22.12	<1	nd	nd	nd	nd	nd	nd
6	26.29	<1	nd	nd	nd	nd	nd	nd
7	30.30	<1	nd	nd	nd	nd	nd	nd
8	35.40	<1	nd	nd	nd	nd	nd	nd
9	39.23	<1	1 (Coxsackie B4)	nd	nd	nd	nd	nd
10	41.47	<1	nd	nd	nd	nd	nd	nd
11	49.12	<1	nd	nd	nd	nd	nd	nd

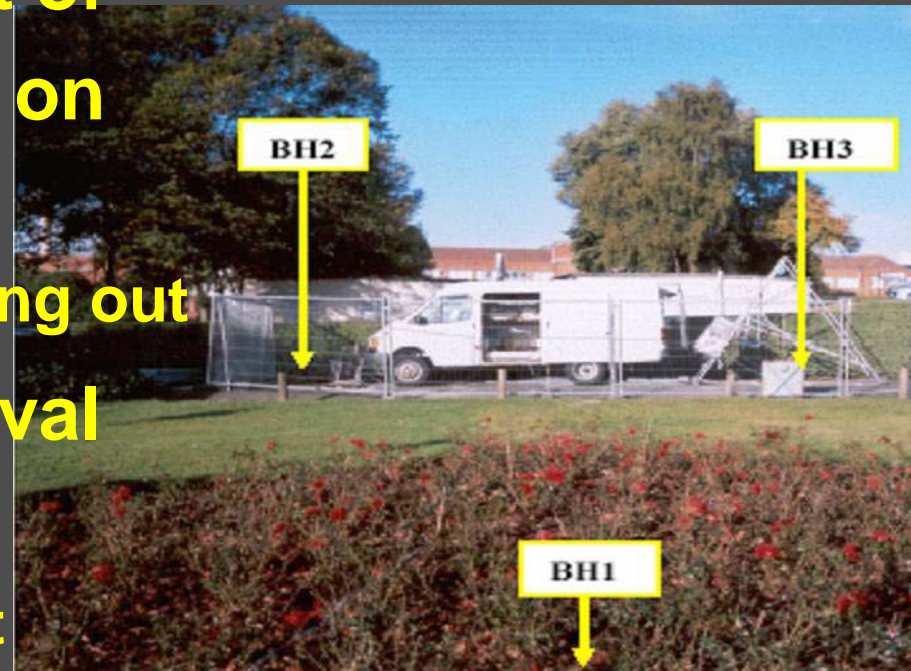
nd - not detected

## ***field monitoring:* aims**

- **to confirm previous finding that viable human viruses exist to at least 40 m depth**
  - **sample example piezometer system**
- **to determine if there is seasonality, and therefore constrain velocity**
  - **sample piezometer monthly for 1.5 years**
- **to determine the frequency of occurrence**
  - **sample wells**

# *field experimentation (phage):* aims

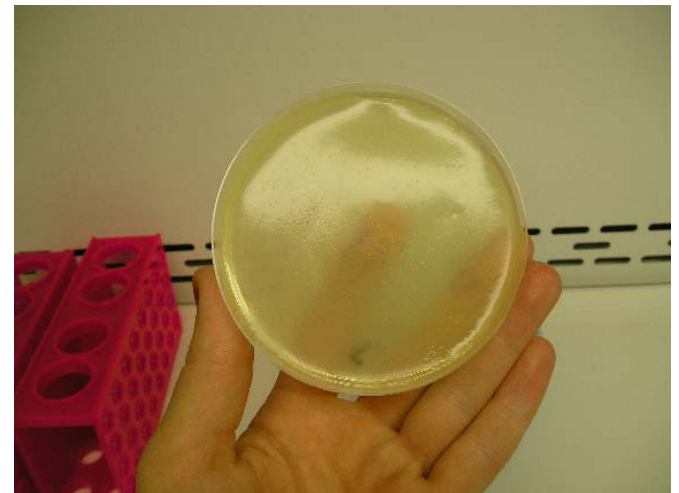
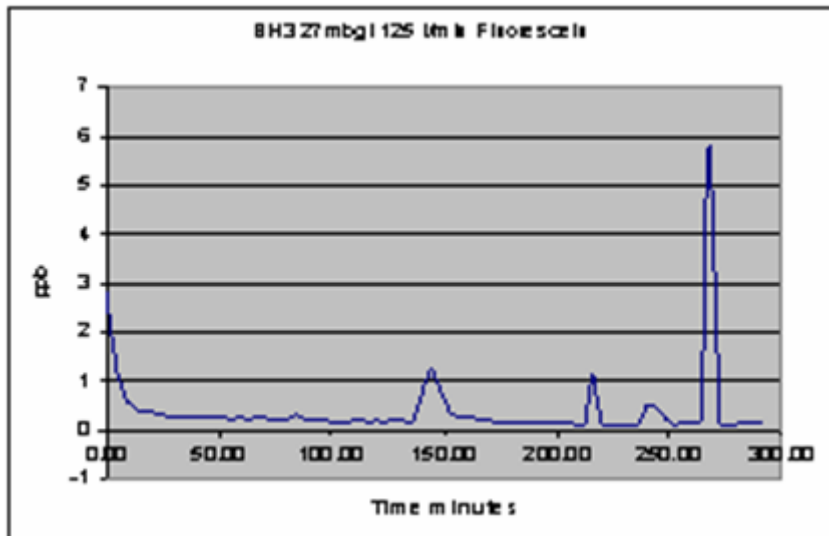
- to determine the effect of residence time on virus attenuation
  - stopped flow push-pull testing
- to determine the effect of velocity / velocity change on virus release
  - vary  $Q$  when pumping out
- to determine the removal of virus with distance
  - undertake 20 m test



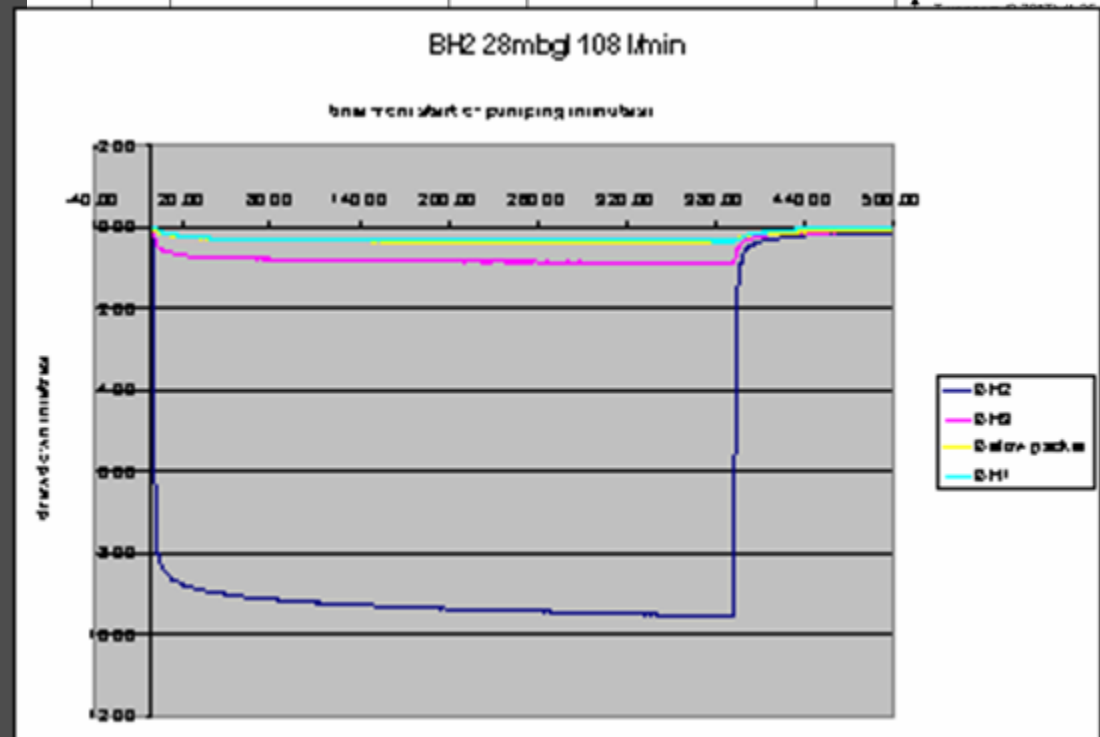
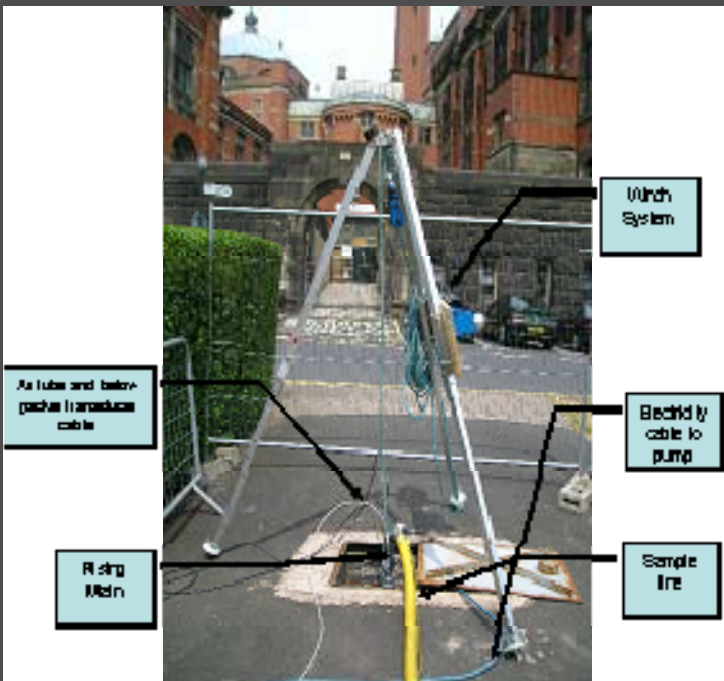
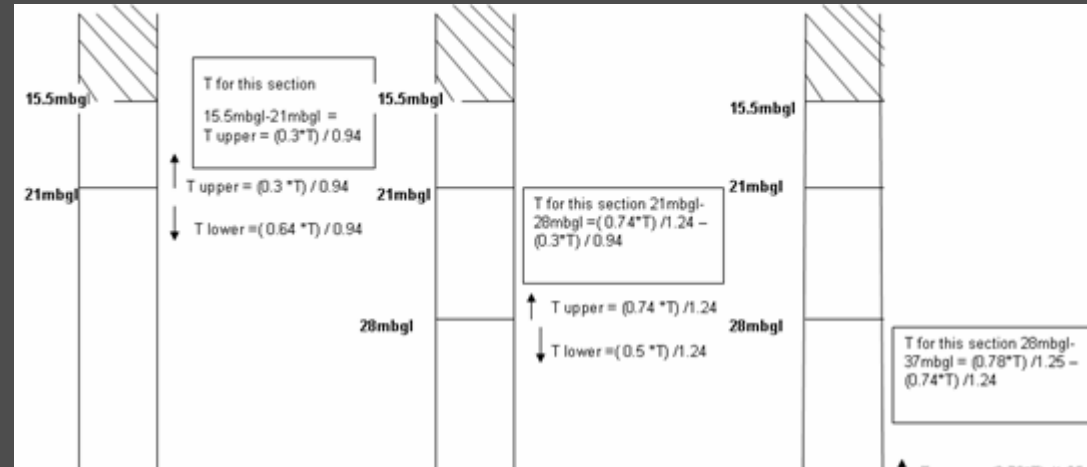


# progress

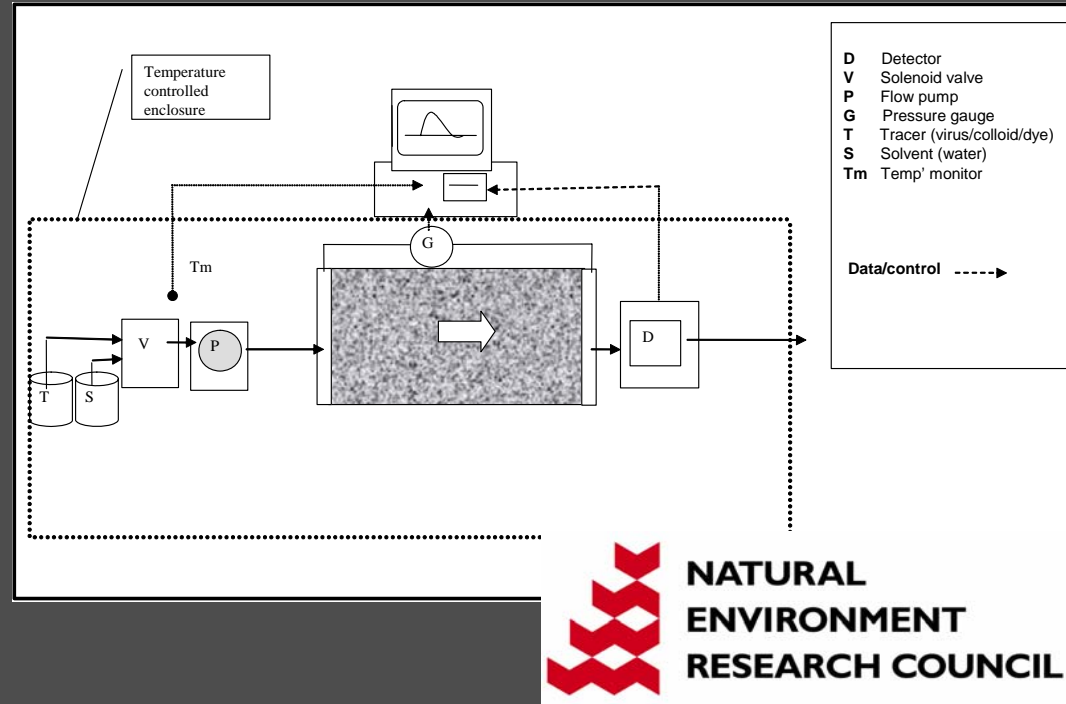
- pumped intervals previously used for injection ~ 1 year earlier
  - no phage detected
  - inactivated, or transported away??



- developed a preliminary flow model of the site



# laboratory experimentation





## ***laboratory experimentation: aims***

- to determine the effect of attachment on the viability of subsequently detached viruses

- stopped-flow expts

- to determine effect of flow velocity on mobility

- use several flow rates

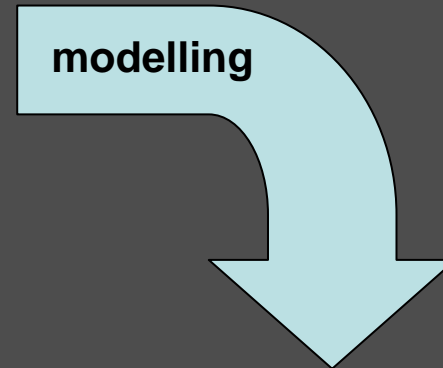
..... if time permits

# Integration

can we use the aquifer's attenuation capacity to make sure that viruses are not present in recovered waste water?

Empirical investigation ....

1. velocities and distances of travel of viable viruses under aquifer conditions
2. effects of (pumping-induced) gw velocity changes
3. effect of attachment on viability



- Hazard?
- Rules to reduce risk





# Integration

## Monitoring

- distance travelled

- velocity in aquifer system

- min pathway frequency

## Field Expts

-  $f(\text{time})$

$f(\text{velocity})$

$f(\text{distance})$

## Lab Expts

-  $f(\text{time})$

-  $f(\text{velocity})$

dependence on  
source  
characteristics

modelling

Hazard?

Rules to reduce  
risk

